## Retinoids and Related Compounds. Part 9.<sup>1</sup> Synthesis and Spectral Characteristics of Retinal Analogues involving the 11-*cis*-Locked-cyclopentatrienylidene Structure

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The 11-*cis*-locked-cyclopentatrienylideneretinals (**7a**), (**7b**), (**8a**), (**8b**), and (**9**) have been obtained in a short-path synthesis from the  $\beta$ -ionyl sulphone (**3**). Their characteristic spectral properties (u.v. and <sup>1</sup>H n.m.r.) are discussed.

The visual pigment rhodopsin (1) has been shown to contain the 11-cis-retinal (2) bound via a protonated Schiff-base linkage to the  $\varepsilon$ -amino group of a specific lysyl residue of the apoprotein opsin.<sup>2</sup> During the course of an investigation<sup>3</sup> into the photobleaching process of rhodopsin, we reported in a previous communication<sup>4</sup> the preparation of the 11-cis-locked<sup>5</sup> cyclopentatrienylidenerhodopsin (7c), whose chromophore consists of the 11-cis- and 12s-trans-fixed retinal analogue (7a). From the photochemical behaviour of the artificial pigment (7c) at low temperatures, the *cis-trans* isomerisation hypothesis for the conversion of rhodopsin into bathorhodopsin was chemically confirmed.<sup>6</sup> A circular dichroism (c.d.) spectrum <sup>6</sup> of (7c), having a non-twisted structure around a 12s-trans bond, showed a negligible  $\alpha$ -band. A comparison of the c.d. data for rhodopsin (1) and cycloheptatrienylidenerhodopsin<sup>7</sup> with our results shows that the presence of the  $\alpha$ -band in the induced c.d. of rhodopsin is due to the twisted 12s-bond in the chromophore. In this paper, we describe the full synthesis and characterisation of the 11-cis-locked cyclopentatrienylideneretinals.

The lithium derivative obtained from the reaction of the  $\beta$ ionyl sulphone (3)<sup>8</sup> with lithium di-isopropylamide (LDA) at -78 °C was treated with 4-acetoxycyclopentenone (4a)<sup>9</sup> in a mixture (ca. 2:1) of dry tetrahydrofuran (THF) and hexa-

methylphosphoric triamide (HMPA) at -50 °C—room temperature to give a mixture of two  $\beta$ -ionylidenecyclopentenone isomers (5a) and (6a) in ca. 24% yield, which could be cleanly separated by a combination of silica gel column chromatography and low-pressure liquid chromatography. The ratio of (5a) to (6a) was ca. 3:2. A trans-configuration was assigned to the newly formed tetrasubstituted 9,10-double bond in the faster eluted isomer (5a), the latter having a u.v. absorption maximum at 350 nm; a comparison with the <sup>1</sup>H n.m.r. data of the other isomer (6a) shows that the 8-H signal of (6a) ( $\delta$  6.66) is deshielded by the 11,12-double bond and that the 11-H signal  $(\delta 8.22)$  appears at lower field owing to the anisotropic effect of the 7,8-double bond. The  $14-H_2$  signals for both isomers appear at  $\delta$  ca. 3 as a singlet, which is characteristic for the position between two sp<sup>2</sup> carbons. This reaction failed both when (4a) was replaced by (4b)<sup>9</sup> and in the absence of HMPA. An Emmons-Horner reaction of the 9-trans-isomer (5a) with diethyl cyanomethylphosphonate using butyl-lithium in dry THF at 0 °C under argon produced a pentaene nitrile which, without purification, was subsequently reduced with diisobutylaluminium hydride (DIBAH) in hexane at -70 °C to furnish a mixture (ca. 4:1) of the 11-mono-cis-aldehyde (7a) and the 11-cis,13-cis-isomer (9) in low yield. Each unstable isomer



		(7 <b>a</b> )	( <b>7b</b> )	( <b>8a</b> )	( <b>8b</b> )	(9)
U.v. (EtOH) λ <sub>max.</sub> nm (ε)		405 (18 500) 263 (13 300)	400 260	402 (11 500)	395	392 (10 000)
<sup>1</sup> H n.m.r. (200 MHz)	1-Me <sub>2</sub>	1.05 (s)	1.03 (s) 1.07 (s)	1.04 (s)	1.05 (s)	1.04 (s)
$(\delta, CDCl_3)$	5-Me	1.76 (s)	1.75 (s)	1.74 (s)	1.74 (s)	1.75 (s)
(J in Hz)	9-Me	2.05 (s)	2.05 (s)	2.02 (s)	2.06 (s)	2.05 (s)
	7-H	6.32 (d, J 16)		6.27 (d, J 16)	6.28 (d, J 16)	
	8-H	6.42 (d, J 16)	50.37 (s-like)	6.62 (d, J 16)	6.60 (d, J 16)	>6.33 (s-like)
	11 <b>-H</b>	7.27 (d, J 5)	7.16 (d, J 5)	7.36 (d, J 5.5)	7.25 (d, J 5)	7.36 (dd, J 1.5, 5.5)
	12 <b>-H</b>	6.61 (d, J 5)	6.49 (d, J 5)	6.59 (d, J 5.5)	6.48 (d, J 5)	7.28 (d, J 5.5)
	14- <b>H</b>	5.97 (td, J 1.5, 7)	5.89 (d, J 8)	5.98 (td, J 2, 7)	5.90 (d, J 8)	5.88 (dd, J 1.5, 8)
	20-H	3.73 (d, J 1.5)	4.04 (q, J 7)	3.70 (d, J 2)	3.98 (q, J 7)	3.48 (s)
	20-Me		1.35 (d, J 7)		1.38 (d, J 7)	
	СНО	9.88 (d, J 7)	9.90 (d, J 8)	9.89 (d, J 7)	9.91 (d, J 8)	9.99 (d, J 8)

Table. U.v. absorption maxima and <sup>1</sup>H n.m.r. data for cyclopentatrienylideneretinals (7a), (7b), (8a), (8b), and (9)

was purified by preparative t.l.c. followed by h.p.l.c. in the dark. The 9-cis-isomer (6a) was treated in the same way as the 9-transisomer (5a) to provide mainly the 9-cis,11-cis-aldehyde (8a), together with (7a) and (9) in low yield, which were carefully separated into their respective isomers by a combination of preparative t.l.c. and h.p.l.c. in the dark. The structures of the three products were confirmed on the basis of their u.v. and <sup>1</sup>H n.m.r. data (Table). The three isomers have absorption maxima at longer wavelengths (ca. 20 nm) than that (380 nm) of the all-trans-retinal compound; this suggests that the former have greater chromophoric coplanarity. This behaviour is particularly notable for the 11-mono-cis-isomer (7a) the u.v. maximum of which is of 30 nm longer wavelength than that (375 nm) of 11-cis-retinal (2). This is the first example of a retinal analogue having the same chromophore as (2), indicating that (7a) has a considerable degree of coplanarity in its conjugated structure. The configuration of the 13,14-double bond was deduced on the basis of the chemical shifts of the 20-H<sub>2</sub> signals. In the 13-trans-isomers (7a) and (8a), C-20 protons were strongly deshielded by the anisotropic effect of the aldehyde group, although the corresponding protons in the 13cis-isomer (9) were not so affected. Instead, 12-H in (9) was observed at low field; this is due to the effect of the aldehyde group. The stereochemistry of the 9,10-double bond was also determined from the chemical shifts of the 8-H signals which were subject to the anisotropic effects of the 11,12-double bond. Thus, the 8-H signal in the 9-cis-isomer (8a) appeared at lower field than the corresponding signals of the 9-trans-isomers (7a) and (9). The <sup>1</sup>H n.m.r. data of the most significant 11mono-cis-isomer (7a) have also been compared with those 10 of all-trans-retinal. The 8-H signal ( $\delta$  6.42) of (7a) is further downfield than the corresponding resonance ( $\delta$  6.18) for the alltrans-retinal, presumably owing to the deshielding anisotropic effect of the C(10)-C(20) single bond. In addition, the 12-H signal ( $\delta$  6.61) appears at lower field than the corresponding signal ( $\delta$  6.37) of all-*trans*-retinal, showing the anisotropic effect of the 13,14-double bond. These spectral data confirm that the 11-cis-locked retinal (7a) has a high degree of coplanarity, particularly in the C(9)-C(14) triene part. This is further supported by MO calculations.<sup>11</sup>

Treatment of a mixture of the two cyclopentenone derivatives (5a) and (6a) with a 10 molar excess of MeI in the presence of lithium bis(trimethylsilyl)amide or LDA yielded the monomethylated compounds (5b) and (6b) which, without separation, were converted into a mixture of the pentaene aldehyde isomers in a similar manner to that used for the 20unmethylated aldehydes (7a), (8a), and (9). The pure isomers (7b) and (8b) were obtained by preparative t.l.c. followed by h.p.l.c. in the dark. The structures of the isomers were deduced from a comparison of their spectral data (Table) with those of the 20-unmethylated aldehydes.

In a binding experiment <sup>6</sup> using bovine opsin, (7b) gave no pigment while (7a) produced the artificial pigment (7c) at a rate which was slow compared with that of the regeneration of rhodopsin. This suggests that, for the regeneration of rhodopsin, selective binding of opsin to a twisted 11-cis-retinal (2) in a 12s-transoid form, and a spatial restriction in an opsin cavity, are strict requirements.

## Experimental

U.v. spectra were recorded on a Shimadzu UV 200S instrument in ethanolic solutions. <sup>1</sup>H N.m.r. spectra at 200 MHz were determined on a Varian XL-200 superconducting FT-NMR spectrometer in deuteriochloroform solutions using tetramethylsilane as an internal reference. The results are given mainly in the Table. Mass spectra were determined on a JEOL JMS-01SG or a Hitachi M-80 double-focusing GC mass spectrometer. Preparative t.l.c. was performed on silica gel plates (Merck silica gel  $60F_{254}$  precoated plates, 0.25 or 0.5 mm thickness). Silica gel used for column chromatography was Merck Kieselgel 60 (particle size 0.063-0.200 mm, 70-230 mesh ASTM). Low-pressure column chromatography was carried out using a Lobar Column (Merck). H.p.l.c. was carried out on a Shimadzu LC-3A instrument with a u.v. detector.

Unless otherwise stated, solvent extracts were dried over anhydrous sodium sulphate and all operations were carried out under nitrogen or argon. Ether refers to diethyl ether.

transand cis-4-[1-Methyl-3-(2,6,6-trimethylcyclohex-1envl)prop-2-envlidene]cvclopent-2-en-1-one (5a) and (6a).—A solution (2.7 ml) of butyl-lithium (15% w/v in hexane) was added to a stirred solution of di-isopropylamine (650 mg) in dry THF (1 ml) at -78 °C, and the mixture was stirred for a further 30 min. To this LDA solution was added a solution of the  $\beta$ ionyl sulphone (3) (2 g) in a mixture of dry THF (1.5 ml) and dry HMPA (1.5 ml) in portions. After the addition was complete, the mixture was stirred at a temperature of -78 °C to -50 °C for ca. 1 h, after which 4-acetoxycyclopent-2-en-1-one (4a) (600 mg) was added dropwise at -50 °C over a period of 30 min. The mixture was then stirred at -30 °C for 1 h before being allowed to warm to room temperature over 2 h with stirring. The reaction was guenched with saturated aqueous NH<sub>4</sub>Cl and extracted with ethyl acetate. The extracts were washed with brine, dried, and evaporated to give an oil which was purified by silica gel column chromatography (5% ether in benzene)

followed by low-pressure column chromatography (LiChroprep Si 60 Merck, hexane-benzene-ether 10:10:1) to afford, as yellow oils, the *trans*-isomer (**5a**) (150 mg), the *cis*-isomer (**6a**) (100 mg), and a mixture of both isomers (15 mg) (yield 24%). The *trans*-isomer (**5a**):  $\lambda_{max}$ . 350 nm ( $\epsilon$  17 500);  $\delta$  1.05 (6 H, s, gemMe), 1.73 (3 H, s, 5-Me), 2.08 (3 H, s, 9-Me), 3.05 (2 H, s, 14-H<sub>2</sub>), 6.24 (1 H, d, J 5.6 Hz, 12-H), 6.30 (1 H, d, J 16 Hz, 8-H), 6.44 (1 H, d, J 16 Hz, 7-H), and 8.14 (1 H, d, J 5.6 Hz, 11-H) (Found:  $M^+$ , 256.184. C<sub>18</sub>H<sub>24</sub>O requires  $M^+$ , 256.183). The *cis*-isomer (**6a**):  $\lambda_{max}$ . 346 nm ( $\epsilon$  14 500);  $\delta$  1.04 (6 H, s, gemMe), 1.74 (3 H, s, 5-Me), 2.00 (3 H, s, 9-Me), 3.03 (2 H, s, 14-H<sub>2</sub>), 6.23 (1 H, d, J 5.6 Hz, 11-H) (Found:  $M^+$ , 256.184. C<sub>18</sub>H<sub>24</sub>O requires  $M^+$ , 256.183).

trans- and cis-5-Methyl-4-[1-methyl-3-(2,6,6-trimethylcyclohex-1-enyl)prop-2-enylidene]cyclopent-2-en-1-one (5b) and (6b).—A solution (0.8 ml) of butyl-lithium (15% w/v in hexane) was added to a stirred solution of hexamethyldisilazane (0.4 ml) in dry 1,2-dimethoxyethane (DME) (0.6 ml). The mixture was boiled under reflux for 0.5 h and cooled to -75 °C. A solution of  $\beta$ -ionylidenecyclopent-2-en-1-one [a mixture of (5a) and (6a)] (273.7 mg) in dry DME (0.6 ml) was added to the above cooled mixture and a solution of MeI (1.52 g) in dry DME (1 ml) was added at -50 °C. The reaction mixture was stirred at -20 °C for 0.5 h, at 0 °C for 1 h, and at room temperature for 1.5 h. The reaction was quenched by the addition of saturated aqueous NH<sub>4</sub>Cl. The ethereal extracts were washed with brine, dried, and evaporated to give an oily mixture (75 mg, 26%) which was separated using a centrifugal thin layer chromatotron (Harrison Model 7924, Merck silica gel 60PF254, 2 mm thickness, hexane-benzene 1:1) followed by h.p.l.c. (LiChrosorb  $0.8 \times 30$  cm, 15% di-isopropyl ether in hexane) to give two pure isomers  $[(5b), \text{longer } R_i; (6b),$ shorter  $R_1$  as yellow oils. This methylation using LDA as a base instead of lithium bis(trimethylsilyl)amide resulted in the same yield. The *trans*-isomer (**5b**):  $\lambda_{max}$ . 350 nm;  $\delta$  1.03, 1.06 (each 3 H, s, gemMe), 1.29 (3 H, d, J 7.6 Hz, 14-Me), 1.74 (3 H, s, 5-Me), 2.08 (3 H, s, 9-Me), 3.00 (1 H, q, J 7.6 Hz, 14-H), 6.15 (1 H, d, J 5.6 Hz, 12-H), 6.33 (1 H, d, J 16 Hz, 8-H), 6.46 (1 H, d, J 16 Hz, 7-H), and 8.11 (1 H, d, J 5.6 Hz, 11-H); m/z 270 ( $M^+$ ). The cis-isomer (**6b**):  $\lambda_{max.}$  346 nm;  $\delta$  1.05 (6 H, s, <sub>gem</sub>Me), 1.32 (3 H, d, J 7.5 Hz, 14-Me), 1.74 (3 H, s, 5-Me), 2.05 (3 H, s, 9-Me), 2.92 (1 H, q, J 7.5 Hz, 14-H), 6.15 (1 H, d, J 5.6 Hz, 12-H), 6.37 (1 H, d, J 15.9 Hz, 7-H), 6.66 (1 H, d, J 15.9 Hz, 8-H), and 8.19 (1 H, d, J 5.6 Hz, 11-H); m/z 270 ( $M^+$ ).

11-cis-, 11-cis,13-cis-, and 9-cis,11-cis- 4-[1-Methyl-3-(2,6,6trimethylcyclohex-1-enyl)prop-2-enylidene]cyclopent-2-enylideneacetaldehyde (**7a**), (**9**), and (**8a**).—Butyl-lithium (90 mg) (15% w/v in hexane; 0.6 ml) was added to a stirred solution of diethyl cyanomethylphosphonate (210 mg) in dry THF (2 ml) and stirring was continued at room temperature for ca. 1 h. This carbanion solution was added to a solution of trans- $\beta$ ionylidenecyclopent-2-en-1-one (**5a**) (230 mg) in dry THF (1 ml) at 0 °C in the dark and stirring was continued for 1 h. The reaction mixture was poured into water and extracted with ether. The ether extract was washed with brine, dried, and evaporated under reduced pressure to give a brown oil which was submitted to preparative t.l.c. (15% ether in benzene) to yield a nitrile (58 mg) which was not further purified. A solution of DIBAH (44 mg) in hexane (1 ml) was added to a solution of the nitrile (58 mg) in hexane (2 ml) at -70 °C in the dark and the mixture was stirred for ca. 10 min. The reaction was quenched by addition of an excess of ether. The ether layer was washed with brine, dried, and evaporated to give a deep orange oil (14 mg) which was a mixture (ca. 4:1) of the 11-cis-isomer (7a) and the 11-cis,13-cis-isomer (9). Preparative t.l.c. (15% ether in hexane) and repeated h.p.l.c. [ $\mu$ -Porasil (0.8  $\times$  30 cm)/hexane-benzene-ether 16:16:1] of the mixture in the dark provided pure (7a) and (9) as orange oils. The 11-cis-isomer (7a):  $\lambda_{\text{max}}$  see Table (Found:  $M^+$ , 282.199. C<sub>20</sub>H<sub>26</sub>O requires  $M^+$ , 282.198). The 11-cis,13-cis-isomer (9):  $\lambda_{max}$  see Table;  $\delta$ , see Table (Found:  $M^+$ , 282.197.  $C_{20}H_{26}O$  requires  $M^+$ , 282.198). 9-cis-\beta-Ionylidenecyclopent-2-en-1-one (6a) (230 mg) was treated in the same way as the 9-trans-isomer (5a) to yield the corresponding nitrile (37 mg) which was converted into the 9-cis,11-cis-isomer (8a) in 25% yield:  $\lambda_{max}$  see Table;  $\delta$ , see Table (Found: M<sup>+</sup>, 282.201. C<sub>20</sub>H<sub>26</sub>O requires M<sup>+</sup>, 282.198).

11-cis- and 9-cis,11-cis- 5-Methyl-4-[1-methyl-3-(2,6,6-trimethylcyclohex-1-enyl)prop-2-enylidene]cyclopent-2-enylideneacetaldehyde (7b) and (8b).—A mixture of (5b) and (6b) (375 mg), treated in a manner similar to that for the preparation of (7a), (9), and (8a) from (5a) and (6a), gave a mixture (47.2 mg) of the 11-cis-aldehyde (7b), the 9-cis,11-cis-aldehyde (8b), and other isomers which were separated and purified by preparative t.l.c. (5% ether in benzene) followed by h.p.l.c. [ $\mu$ -Porasil (0.8 × 30 cm)/hexane-benzene-ether 16:16:1] to afford (7b) and (8b) as orange oils. The 11-cis-aldehyde (7b):  $\lambda_{max}$  see Table;  $\delta$ , see Table (Found:  $M^+$ , 296.214. C<sub>21</sub>H<sub>28</sub>O requires  $M^+$ , 296.214). The 9-cis,11-cis-aldehyde (8b):  $\lambda_{max}$  see Table;  $\delta$ , see Table (Found:  $M^+$ , 296.213. C<sub>21</sub>H<sub>28</sub>O requires  $M^+$ , 296.214).

## References

- 1 Part 8, M. Ito, T. Iwata, K. Tsukida, Y. Shichida, and T. Yoshizawa, J. Nutr. Sci. Vitaminol., 1984, 30, 577.
- 2 (a) 'Methods in Enzymology,' ed. L. Packer, Academic Press, New York, 1982, 81; 1982, 88; (b) Y. A. Ovchinnikov, FEBS Lett., 1982, 148, 179.
- 3 (a) M. Ito, K. Hirata, A. Kodama, K. Tsukida, H. Matsumoto, K. Horiuchi, and T. Yoshizawa, Chem. Pharm. Bull., 1978, 26, 925; (b) S. Kawamura, T. Yoshizawa, K. Horiuchi, M. Ito, A. Kodama, and K. Tsukida, Biochim. Biophys. Acta, 1979, 548, 147; (c) M. Ito, A. Kodama, M. Murata, M. Kobayashi, K. Tsukida, Y. Shichida, and T. Yoshizawa, J. Nutr. Sci. Vitaminol., 1979, 25, 343; (d) Y. Fukada, T. Yoshizawa, M. Ito, and K. Tsukida, Biochim. Biophys. Acta, 1982, 708, 112.
- 4 M. Ito, A. Kodama, K. Tsukida, Y Fukada, Y. Shichida, and T. Yoshizawa, *Chem. Pharm. Bull.*, 1982, **30**, 1913.
- 5 Retinoid numbering according to the recommendations of the IUPAC-IUB Joint Commission on Biochemical Nomenclature (J. Biol. Chem., 1983, **258**, 5329) was employed in the formulae.
- 6 Y. Fukada, Y. Shichida, T. Yoshizawa, M. Ito, A. Kodama, and K. Tsukida, *Biochemistry*, 1984, 23, 5826.
- 7 H. Akita, S. P. Tanis, M. Adams, V. Balogh-Nair, and K. Nakanishi, J. Am. Chem. Soc., 1980, 102, 6370.
- 8 A. Fischli, H. Mayer, W. Simon, and H.-J. Stoller, *Helv. Chim. Acta*, 1976, **59**, 397.
- 9 C. H. DePuy, M. Isaks, K. L. Eilers, and G. F. Morris, J. Org. Chem., 1964, 29, 3503.
- 10 R. S. H. Liu and A. E. Asato, Tetrahedron, 1984, 40, 1931.
- 11 Y. Beppu, T. Kakitani, M. Ito, K. Tsukida, and T. Yoshizawa, Chem. Phys. Lett., 1985, 116, 231.

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